

Biodistribution of sodium pertechnetate and biochemical parameters in experimental osteoporosis in rats treated with extract of *Chenopodium ambrosioides* L (mastruz)

Biodistribuição do pertecnetato de sódio e parâmetros bioquímicos em modelo experimental de osteoporose em ratas tratadas com extrato de *Chenopodium ambrosioides* L (mastruz)

Ciro Dantas Soares, Sérgio Rodrigo Pereira Trindade, Tarciso Bruno Montenegro Sampaio, Cipriano Galvão da Trindade Neto, Rejane Andrade de Carvalho, Maria Goretti Freire de Carvalho, Márcia Martins Marques, Amália Cíntia Meneses Rêgo, Irami Araújo-Filho, Aldo Cunha Medeiros.

Performed at Coordination of Research and Postgraduate, Potiguar University, Natal, Brazil; and Department of Surgery, Federal University of Rio Grande do Norte (UFRN), Brazil.

Financial support: none.

Conflict of interest: none

Correspondence address: Aldo Cunha Medeiros, Department of Surgery, Federal University of Rio Grande do Norte, at Av. Nilo Peçanha 620, Natal, RN, Brazil, Email: aldo@ufrnet.br

Submitted: 20 Jul 2014. Accepted, after review: 10 Aug 2014.

ABSTRACT

Objective: The aim of this study was to evaluate the effect of treatment with *Chenopodium ambrosioides* L (mastruz) hydroalcoholic extract in rats with osteoporosis on the biodistribution of sodium pertechnetate ($\text{Na}^{99\text{m}}\text{TcO}_4$) and biochemical parameters. **Methods:** Ten rats underwent ovariectomy to induce osteoporosis. The first group ($n = 5$) received no treatment, the second group ($n = 5$) received 50mg HaME for thirty days. After finishing the treatment the animals had been anesthetized and injected 0.1 mL (0.66 MBq) of $\text{Na}^{99\text{m}}\text{TcO}_4$ the femoral vein. After 30 minutes, blood was collected for dosages, and then the animals were killed by an overdose of anesthetic (thiopental 100mg/kg) and removed the heart, lungs, stomach, femur, kidneys, liver, intestine and thyroid to calculate the percentage of radioactivity per gram of tissue (% ATI/ g) of each organ. **Results:** There was no significant difference in any of the organs analyzed, but in all of them, except in the intestine, the uptake of $\text{Na}^{99\text{m}}\text{TcO}_4$ was higher in the group treated with mastruz. Biochemical parameters showed a significant increase in lactate dehydrogenase, total cholesterol, and aspartate aminotransferase. These results allow us to state that mastruz may have hepatotoxic effect. **Conclusion:** Treatment with mastruz in rats with osteoporosis does not alter the biodistribution of the radiopharmaceutical $\text{Na}^{99\text{m}}\text{TcO}_4$ and showed some hepatotoxicity

Key words: Osteoporosis. *Chenopodium ambrosioides* L. Bioavailability. $^{99\text{m}}$ Tc. Pertechnetate. Rats.

RESUMO

Objetivo: O objetivo deste estudo foi avaliar o efeito do tratamento com extrato hidroalcoólico de *Chenopodium ambrosioides L* (mastruz) em ratas com osteoporose na biodistribuição do pertecnetato de sódio ($\text{Na}^{99\text{m}}\text{TcO}_4$) e em parâmetros bioquímicos.

Métodos: Dez ratas foram submetidas à ooforectomia para indução de osteoporose. O primeiro grupo (n=5) não recebeu tratamento, e o segundo grupo (n=5) recebeu diariamente 50 mg de EhaM por trinta dias. Depois de finalizado o tratamento, os animais foram anestesiados e injetado 0,1 mL (0,66MBq) de $\text{Na}^{99\text{m}}\text{TcO}_4$ através da veia femural. Após 30 minutos foi coletado sangue e em seguida os animais foram mortos com superdose de anestésico (tiopental 100mg/Kg) e removidos coração, pulmões, estômago, fêmur, rins, fígado, intestino e tireóide para cálculo do percentual de radioatividade por grama de tecido (%ATI/g) de cada órgão. **Resultados:** Não houve diferença significativa em nenhum dos órgãos analisados, porém em todos eles, exceto no intestino, a captação do $\text{Na}^{99\text{m}}\text{TcO}_4$ foi maior no grupo tratado com mastruz. Dos parâmetros bioquímicos analisados houve aumento significativo de Lactato desidrogenase, colesterol total e aspartato aminotransferase. Esses resultados permitem afirmar que as provas de função hepática sinalizam que o mastruz pode apresentar efeito hepatotóxico. **Conclusão:** O tratamento com mastruz em ratas com osteoporose não altera a biodistribuição de $\text{Na}^{99\text{m}}\text{TcO}_4$, mas pode apresentar hepatotoxicidade nas doses empregadas.

Descritores: Osteoporose. *Chenopodium ambrosioides L*. Biodisponibilidade. Pertecnetato. Tecnécio $\text{Tc}^{99\text{m}}$. Ratos.

INTRODUCTION

Diagnostic techniques have evolved in recent decades and among the advances are the advent of bone scintigraphy, a relevant technique for detection of bone metastases, primary tumors and benign bone diseases¹⁻³. However, several pathological conditions can alter the biodistribution of radiopharmaceuticals used in these exams^{4,5}. We did not find studies assessing the biodistribution of radiopharmaceuticals in rats treated with mastruz. The sodium pertechnetate ($\text{Na}^{99\text{m}}\text{TcO}_4$) is one of the most widely used isotope in nuclear medicine and research on their biodistribution in various physiological and pathological situations have been done^{2,3}.

Osteoporosis (OP) is a metabolic bone disease, considered a public health problem as it affects at least 30% of women in post-menopause. It is a chronic disease caused by increased osteoclastic activity and suppression of osteoblastic activity. Clinically various effects of this disease, including increased risk of bone fractures,⁴⁻⁶ are observed. The mastruz has osteogenic effect, evidenced by Souza et al (2010) and may be a promising therapy for osteoporosis and other bone diseases⁷.

The *Chenopodium ambrosioides L* (mastruz) is a medicinal plant widely used in the world. Studies have proved their anthelmintic, anti-inflammatory and analgesic

activities. It has osteogenic effect and hence its effect has been studied in osteoporosis. The main advantages of using herbal medicines are easy accessibility (cost and availability) and reduction of adverse effects compared with conventional drugs⁷⁻⁹.

Systemic effects of herbal medicines are essential for indicating or not for the treatment of diseases. Some drugs (biphosphonates) used to treat osteoporosis are associated with alterations in biodistribution of radiopharmaceuticals, and may result in misinterpretation of images in nuclear medicine^{6,10}. Capriles *et al.* evaluated the influence of eggplant extract on the biodistribution of technetium in blood of rats¹¹.

The aim of this study was to evaluate the biodistribution of sodium pertechnetate ($\text{Na}^{99\text{m}}\text{TcO}_4$) and biochemical parameters in rats with induced osteoporosis by bilateral oophorectomy, treated with mastruz hydroalcoholic extract (MaHaE).

METHODS

The experimental protocol was submitted to the Ethics Committee (CEP/UNP) and approved under number 007/2013. This protocol followed the recommendations of the Animal Experimentation Code of Ethics and Brazilian College of Animal Experimentation.

Extract

Fresh plant *Chenopodium ambrosioides L* was collected and placed in drying oven for three days. Then ethanol was added and homogenized in 1:3 ratio for percolation process. The material was filtered and concentrated in rotoevaporator at constant temperature of 60°C. The concentrated extract was weighed and diluted in distilled water, obtaining hydroalcoholic mastruz extract (MaHaE) at a concentration of 10%, maintained under refrigeration (5°C). All experiments were carried out before the expiry date of the product.

Surgery and sample collection

Ten female Wistar rats 90 days old, 200g ± 50g, provided by the Animal Facility of Potiguar University were kept in an environment with lighting (cycles of 12 h light / dark), Ventilation and appropriate temperature (21°C), getting balanced diet (Labina® Purina) and water *ad libitum*.

Bilateral ovariectomy was performed for induction of osteoporosis in all animals. After surgery the rats were divided into two groups: Osteoporosis (G1, n = 5) received 0.5 ml distilled water by gavage for 30 days (G2, n = 5) and osteoporosis+ MaHaE group rats (n = 5), which received daily MaHaE 10% (50 mg) 0.5 mL by gavage at the same route and period.

To perform bilateral ovariectomy, animals were anesthetized using solution of Zoletil 50 at a dose of 0.3 ml/100 mg intramuscular in the quadriceps region. After anesthesia, the anterior abdominal region was shaved, and antisepsis with 2%

chlorhexidine, followed by longitudinal incision with a length of approximately 3 cm in the abdominal wall with exposure of uterus and ovaries. After identification of ovaries, bilateral ligation of mesovaries were done with 4-0 silk sutures (Ethicon® / Johnson & Johnson) and subsequent bilateral oophorectomy with hemostasis review. At the end of the procedure, abdominal wall was closed with Polydioxanone - 4-0 PDS (Ethicon® /Johnson & Johnson) suture and the skin with 4-0 nylon (Ethicon® / Johnson & Johnson). Postoperative pain was treated with tenoxicam 0.5 mg/kg (IM), (Roche® Pharm Brazil) once a day for 3 days. Both groups remained in postoperative observation for 10 days, during which, the rats were daily weighted.

Treatment with distilled water (G1) and matruz extract (MaHaE) (G2) was restarted 24 hours after bilateral oophorectomy.

Biodistribution analysis

Thirty days after the beginning of treatments, animals were anaesthetized again and 0.1 ml of $\text{Na}^{99\text{m}}\text{TcO}_4$ was injected through the femoral vein, corresponding to a radioactivity of 0.66MBq. Technetium-99m was eluted from a 99 Mo/99m Tc generator produced by the Institute of Nuclear Energy Research, São Paulo/Brazil and kindly donated by Northeriogrاندense League against Cancer, Natal / RN. After 30 minutes, blood was collected, then the rats were killed with anesthesia overdose (Tiopental 100mg/Kg i.p.) and organs samples were collected from heart, right lung, stomach, right femur, right kidney, liver, duodenum and thyroid. They were washed in 0.9% saline solution and weighed in a digital precision balance (160-Bel -Mark Itália®-II). The radioactivity uptake of each organ was determined by an automated gamma counter (1470 Wizard, Perkin Elmer®, Finland) with automatic correction for decay and efficiency of 86%, at the Nucleus for Experimental Surgery-UFRN. The counts per minute (CPM) in each organ alone were performed in standard time of 1min. We calculated the percentage of radioactivity per gram of tissue (%ATI/g) in the organs, by dividing the radioactivity present in each organ by the total radioactivity injected into the animal.

Laboratory analysis

Biochemical parameters were measured using the auto-analyzer spectrophotometer (Konelab 60i®) in the Laboratory of Clinical Analysis of Federal University of Rio Grande do Norte, Brazil. Serum lactate dehydrogenase (LDH), aspartate aminotransferase (AST), cholesterol, glucose, triglycerides and globulin were measured using test kits Weiner® São Paulo, Brazil. All data are presented as mean±standard deviation.

Statistical analysis

The data of %ATI/g were analysed in GraphPad PRISM® 5.0 software and statistically processed using the Mann-Whitney test and Student t test. Data were considered significant at a level of 5% ($p < 0.05$).

RESULTS

Biodistribution of Na^{99m}TcO₄

The results of the Na^{99m}TcO₄ uptake in different organs selected for this study are summarized in Table 1.

Table 1 – Percent of radioactivity per gram (%ATI/g) of tissue in each organ.

Organs	%ATI/g		p-value
	Osteoporosis	Osteoporosis+ MaHaE	
Liver	0.18 ± 0.0112	0.25 ± 0.0443	0.1447
Intestine	0.44 ± 0.0861	0.31 ± 0.0506	0.2427
Heart	0.12 ± 0.0111	0.14 ± 0.0139	0.2946
Kidney	0.33 ± 0.0321	0.34 ± 0.0241	0.8099
Thyroid	4.81 ± 0.786	5.51 ± 1.252	0.6495
Lung	0.26 ± 0.0212	0.31 ± 0.0312	0.2391
Femur	0.12 ± 0.0096	0.17 ± 0.0566	0.4093
Estomach	2.73 ± 0.3559	3.55 ± 0.2636	0.1026

*Mean±standard deviation. No significant difference comparing the groups (p<0.05).

The values of the percentage of the Na^{99m}TcO₄ activity in all organs showed no statistically significant differences when the two groups were compared (p>0.05).

Biochemical parameters

Serum levels of AST, LDH, total cholesterol, triglycerides and globulin were significantly higher in the group treated with mastruz than in the control group (p<0.03). The levels of glucose in the blood of animals treated with mastruz were lower than in controls, but the difference was not statistically significant (p=0.0727). These data are summarized in table 2.

Table 2 – Effect of 30 days mastruz (HaME) treatment on biochemical parameters.

Parameters	Osteoporosis	Osteoporosis + HaME	p-value
Glucose	147.0 ± 13.63	110.2 ± 12.31	0.072
AST	132.2 ± 15.61	246.2 ± 33.19	0.011*
LDH	2539 ± 754.3	602.7 ± 36.42	0.028*
Cholesterol	67.50 ± 5.340	84.67 ± 4.088	0.028*
Triglycerides	24.33 ± 1.764	57.17 ± 2.455	< 0.0001*
Globulin	4.470 ± 0.2720	1.783 ± 0.1869	< 0.0001*

Mean±standard deviation. * Statistically significant differences comparing the groups (p<0,05); HaME, Hidro-alcoholic mastruz extract.

DISCUSSION

The mastruz is one of the most widely used herbal medicines in the world. Several medicinal properties have been attributed to mastruz and its use is common in

many regions of Brazil. Recently, mastruz was included by the Brazilian government in the list of priorities for study on the National Program on Medicinal Plants and Herbal Medicines¹⁴. Amounts of flavonoids, tannins and phenols are found in phytochemical analysis of the plant. The main component of the essential oil that can also be found in the crude extract of the plant is ascaridol. The ascaridol and their isomeric forms are organic components of the monoterpenes group, with antioxidant, analgesic, anthelmintic and sedative properties^{9,12-14}. Souza et al. observed osteogenesis inducing activity promoted by mastruz and so it may be a promising candidate for treatment of bone disorders including osteoporosis¹⁵. However, despite the beneficial pharmacological effects of monoterpenes present in the mastruz extract, its chronic use may be toxic to certain organs. Further research are needed in this area of knowledge^{13,15}.

In this study there was a significant increase in AST, LDH and cholesterol, suggesting a possible hepatotoxic effect. Other studies evaluated the effects of chronic administration of mastruz. In the present essay, to evaluate the therapeutic effect of mastruz in osteoporosis, we used 50mg daily for 30 days. It should be noted that the animals in this study were treated with mastruz hydroalcoholic extract, then the amount of alcohol may have induced these alterations. Extracts obtained with the freeze-dried or aired in emulsions plant material can answer these questions and optimize the use of the compounds of mastruz.

Pereira et al. studied administration of mastruz at concentrations of 5, 50 and 500mg/kg by 15 days and concluded that the treatment induced specific changes in the groups treated with high doses, but they were not lethal¹³. They did not use hydroalcoholic extract. Mastruz was manipulated after lyophilization of the plant material, supporting the hypothesis that the hepatotoxic effect may have been from alcohol and not mastruz.

Similar results were described by Holland et al., who treated rats for 10 days with propolis extract, at a dose of 100mg/Kg. They concluded that the treatment altered liver enzymes and alkaline phosphatase, causing a possible hepatotoxic effect¹⁶. The hepatotoxic effects of medicinal plants has been widely studied¹⁷. People need to be alerted that the indiscriminate use of such substances may cause adverse effects. Studies comparing different doses and periods of administration are needed to clarify these questions.

The biodistribution of $\text{Na}^{99\text{m}}\text{TcO}_4$ has an important clinical significance in scintigraphic examinations, in which tissues capture the radiopharmaceutical to produce images. Several physiological, pathological and therapeutic conditions have been associated with changes in the activity $\text{Na}^{99\text{m}}\text{TcO}_4$ such as bariatric surgery, regular administration of herbal medicines, intestinal resection, inflammation, etc^{11,18,19}. The mastruz was not able to alter the biodistribution in organs studied. So, we can infer that in animals and probably in humans, the chronic use of mastruz do not alter the results of scintigraphy with $\text{Na}^{99\text{m}}\text{TcO}_4$, avoiding false negative results. Further studies must be conducted on this line of research to elucidate the unclear questions.

CONCLUSION

According to the experimental model used in this study, it is concluded that administration of mastruz in rats did not alter the biodistribution of sodium pertechnetate. However, elevated levels of liver enzymes, suggest a possible hepatotoxic effect in the period and dose used.

REFERENCES

1. Light TR. Clinical applications of musculoskeletal nuclear medicine. In: Henkin RE, Bova D, Dillehay G, editors. Nuclear Medicine. 2nd ed. Philadelphia: Mosby Elsevier; 2006. p.1108-9.
2. Arano Y. Recent advances in 99mTc radiopharmaceuticals. Ann Nucl Med. 2002; 16:79-93.
3. Ballinger JR. The influence of carrier on 99mTc radiopharmaceuticals. Quarterly J Nucl Med. 2002;46:224-32.
4. Gennari L, Becherini L, Falchetti A, Masi L, Massart F, Brandi ML. Genetics of osteoporosis: role of steroid hormone receptor gene polymorphisms. J Steroid Biochem Mol Biol. 2002;81:1-24.
5. World Health Organ Tech Rep Ser. Prevention and management of osteoporosis. 2003;921:1-164.
6. Miller RG, Chretien KC, Meoni LA, Liu YP, Klag MJ, Levine MA. Comparison of intravenous pamidronate to standard therapy for osteoporosis. J Clin Rheumatol. 2005;11:2-7.
7. Cavalli JF, Tomi F, Bernardini AF, Casanova J. Combined analysis of the essential oil of *Chenopodium ambrosioides* by GC, GC-MS and ¹³C-NMR spectroscopy: quantitative determination of ascaridole, a heat-sensitive compound. Phytochem Anal. 2004;15:275-9.
8. Hallal A, Benali S, Markouk M, Bekkouche K, Larhsini M, Chait A, Romane A, Abbad A, El Abdouni MK. Evaluation of the analgesic and antipyretic activities of *Chenopodium ambrosioides* L. Asian J Biol Sci. 2010;1:894-7.
9. Ibranke GF, Ajiboye KI. Studies on the anti-inflammatory and analgesic properties of *Chenopodium ambrosioides* leaf extract in rats. Int J Pharmacol. 2007;3:111-5.
10. Santos-Oliveira R, Smith SW, Carneiro-Leão AMA. Radiopharmaceuticals drug interactions: a critical review. Ann Acad Bras Cienc. 2008;80:665-75.
11. Capriles PV, Dias AP, Costa TE, Oliveira MB, Faria MV, Moura EG, Abreu BA, Bernardo-Filho M. Effect of eggplant (*Solanum melongena*) extract on the in vitro labeling of blood elements with technetium-99m and on the biodistribution of sodium pertechnetate in rats. Cell Mol Biol. 2002;48:771-6.
12. TrivellatoGrassi L, Malheiros A, Meyre-Silva C, Buss Zda S, Monguilhott ED, Fröde TS, da Silva KA, de Souza MM. From popular use to pharmacological validation: a study of the anti-inflammatory, anti-nociceptive and healing effects of *Chenopodium ambrosioides* extract. J Ethnopharmacol. 2013 Jan 9;145(1):127-38.
13. Pereira WS, Ribeiro BP, Sousa AI, Serra IC, Mattar NS, Fortes TS, Reis AS, Silva LA, Barroqueiro ES, Guerra RN, Nascimento FR. Evaluation of the subchronic toxicity

of oral treatment with *Chenopodium ambrosioides* in mice. *J Ethnopharmacol.* 2010;127:602-5.

14. Brasil. Ministério da Saúde, Secretaria de Ciências, Tecnologia e Insumos Estratégicos. Portaria Interministerial nº 2960 em 9 de dezembro de 2008. Programa de Nacional de Plantas Mediciniais e Fitoterápicos. Diário Oficial da União, Brasília (DF); 2008 Dez 10. Disponível em: http://portal.mda.gov.br/portal/saf/arquivos/view/Programa_Nacional_de_Plantas_Mediciniais_e_Fitoter%C3%A1picos.pdf

15. Souza JNL. Ação da aroeira (*Schinus terebinthifolius* Raddi) e do mastruz (*Chenopodium ambrosioides* L) no processo de reparo de dentes reimplantados de ratos. [Dissertação]. Natal: Universidade Potiguar; 2010.

16. Holanda CM, Barbosa DA, Demeda VF, Bandeira FT, de Medeiros HC, Pereira KR, Barbosa VS, Medeiros AC. Influence of *Annona muricata* (soursop) on biodistribution of radiopharmaceuticals in rats. *Acta Cir Bras.* 2014;29:145-50.

17. Pittler MH, Ernest E. Systematic review: hepatotoxic events associated with herbal products. *Alim Pharmacol Ther.* 2003;18:451-71.

18. Rêgo AC, Ramalho RA, Egito ES, Araújo-Filho I, Azevedo IM, Palestro CJ, Medeiros AC. Biodistribution of technetium-99m pertechnetate after total colectomy in rats. *Appl Radiat Isot.* 2010;68:2169-73.

19. Rêgo AC, Araújo-Filho I, Azevedo IM, Jácome DT, Ramalho Rde A, Medeiros AC. Biodistribution of technetium-99m pertechnetate after Roux-en-Y gastric bypass (Capella technique) in rats. *Acta Cir Bras.* 2010;25:9-12.