Efeito do antimalárico cloroquina na biodistribuição do pertecnetato de sódio em camundongos swiss

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ABSTRACT

Background/Objective: Chloroquine is the prototype anti malarial drug, most widely used to treat all types of malaria except for disease caused by chloroquine resistant Plasmodium falciparum. It is also indicated in the treatment of rheumatoid arthritis, lupus erythematosus, giardiasis, amoebic and autoimmune hepatitis. Technetium-99m (^{99m}Tc) in the form of sodium pertechnetate (Na^{99m}TcO₄) is a radionuclide that connects to wide variety of molecules and cells. This study aimed to evaluate in vivo the influence of chloroquine on the biodistribution of Na^{99m}TcO₄⁻ and histological pattern of Swiss mice. Methods: We used 14 male Swiss mice: treated group (n = 7) received chloroquine solution and control groups (n=7) received filtered water, for 15 days. Both groups received 0.1 mL (0.66 MBg) of Na^{99m}TcO₄⁻ and the percentage of radioactivity per gram of tissue (%ATI/g) was determined. Parts of tissue samples were isolated for histologycal analysis. Results: Statistically significant difference in biodistribution of sodium pertechnetate (ATI%/g) was observed in blood and liver with increased radiotracer uptake in the treated group. Significant histological changes were seen in mesangial cells in the kidney and significant vascular congestion of the treated group. **Conclusion:** We conclude that the use of chloroquine in swiss mice interferes with the uptake of sodium pertechnetate in some organs and histological changes were visualized probably due to the presence of chloroquine toxicity.

Key-words: Chloroquine.Pertechnetate-Tc99m.Radiopharmaceuticals.Bioavailability. Toxicity. Mice.

RESUMO

Introdução/Objetivo: A cloroquina é o protótipo de fármaco antimalárico mais amplamente utilizado para tratar todos os tipos de malária, exceto para a doença causada por Plasmodium falciparum resistente à cloroquina. Também é indicado no tratamento de artrite reumatóide, lúpus eritematoso, giardíase, amebas e hepatite auto-imune. Os radiofármacos são compostos radioativos amplamente utilizados para o diagnóstico e tratamento de várias doenças. Tecnécio-99m (99mTc), sob a forma de pertecnetato de sódio (Na^{99m}TcO₄) é um radionuclídeo que se liga a uma grande variedade de moléculas e células, sendo utilizado em cintilografias do estômago, glândulas salivares, da tiróide e glândulas paratiróides, o plexo coróide, em análises de refluxo esofágico e de fluxo sanguíneo. Este estudo teve como objetivo avaliar in vivo a influência da cloroquina na biodistribuição do Na^{99m}TcO₄ e padrão histológico de camundongos Swiss. Métodos: Foram utilizados 14 camundongos Swiss machos: grupo tratado (n = 7) recebeu solução de cloroquina e controle (n = 7) recebeu água filtrada, por 15 dias. Ambos os grupos receberam 0,1 ml (0,66 MBq) de Na^{99m}TcO₄⁻ eo percentual de radioatividade por grama de tecido (% ATI / g) foi determinada. Amostras de tecidos foram isoladas para análise histológica. Resultados: Foi observada diferença estatisticamente significante na biodistribuição do pertecnetato de sódio (ATI% / g) no sangue e no fígado, com aumento da captação do radiofármaco no grupo tratado. Alterações histológicas significativas foram observadas em células mesangiais do rim e congestão vascular do grupo tratado. Conclusão: Conclui-se que o uso de cloroquina em camundongos swiss interfere na absorção de pertecnetato de sódio em alguns órgãos e alterações histológicas foram visualizadas provavelmente devido à toxicidade da cloroquina.

Descritores: Cloroquina, Pertecnetato de sódio, Biodisponibilidade, radiofármaco, toxicidade, camundongo.

INTRODUCTION

Chloroquine is a4-amino-quinoline considered the prototype anti malarial drug, most widely used to treat all types of malaria except for disease caused by chloroquine resistant *Plasmodium falciparum*. It is highly effective against erythrocytic forms of *Plasmodium vivax*, *Plasmodium ovale* and *Plasmodium malariae*, sensitive strains of *Plasmodium falciparum* and gametocytes of *Plasmodium vivax*¹. Although it has limited action onsome strains of *P.falciparum*, is an important antimalarial been widely used since the 50s due to its efficacy, tolerability, and low cost²⁻³. It is also indicated in the treatment of rheumatoid arthritis, lupus erythematosus, giardiasis and hepatic amoebiasis^{4,5,6,7}. This drug presents systemic adverse reactions affecting the gastrointestinal tract, nervous system, skeletal and cardiac muscle, skin andeyes^{8,9}, accumulates in intracellular acidic organelles, raises the pH, and induces osmotic

swelling and permeabilization of acidic organelles, which account for cloroquineinduced cytotoxicity¹⁰.

After ingestion, chloroquine is rapidly and almost completely absorbed from the gastrointestinal tract(2 –4h)and is deposited in the tissues in considerable quantities in experimental animals, can be found in liver, spleen, kidney and lung from 200to 700 times the plasma concentration¹¹. Leukocytes also concentrate the drug. The brain and spinal cord, moreover, contain only 10 to 30 times the amount present in plasma. After administration, is biotransformed via cytochrome P450 enzymes forming the desetilchloroquine which is pharmacologically active and bisdesetilchloroquine, which are cleaned with a half-life of 20 to 60 days. The drug and metabolitec an be detected in urine months after a single dose¹².

Radiopharmaceuticals are radioactive compounds widely used for diagnosis and treatment of various diseases. Technetium-99m (^{99m}Tc) in the form of sodium pertechnetate (Na^{99m}TcO₄⁻) is a radionuclide that connects to wide variety of molecules and cells widely used radionuclide scintigraphy in the stomach, salivary gland, thyroid and parathyroid glands, the choroid plexus, esophageal reflux and blood flow^{13,14}, in addition to its use in experimental research. The evidence that natural and/or synthetic drugs can affect the biodistribution of radiopharmaceuticals (radiobiocomplexes) in setting of nuclear medicine clinic is already known^{14,15,16}. Several drugs can change the biological effect of the radiopharmaceutical and their interaction can lead to hypo or hyper uptake of radiopharmaceuticals in a particular organ, causing incorrect diagnosis or misinterpretation of results. Repeated scintigraphy may result in unnecessary radiation for patients^{16,17}.

This study aimed to evaluate in vivo the influence of chloroquine on the biodistribution of $Na^{99m}TcO_4^-$ and histological pattern in organs of Swiss mice.

METHODS

We used 14 male Swiss mice weighing 20±5 g from the vivarium of the Potiguar University of Rio Grande do Norte, Natal, Brazil. The animals had free access to water and standard rodent food (Purina/Labina®) maintained under constant environmental conditions (23±2°C; 12h/12h of light/dark cycle). The protocol was conducted according to international regulations for animal experimentation and approved by the Research Ethics Committee of Bioscience Center of the Federal University of Rio Grande do Norte (UFRN) with the protocol number CEUA/058/2011. The Na^{99m}TcO₄⁻ was eluted from a ⁹⁹Mo/^{99m}Tc generator produced by the Institute of Energy and Nuclear Research, São Paulo/Brazil and kindly supplied by the Norteriograndense League

Against Cancer. The animals were randomly allocated to two groups of seven animals each one: treated group (n = 7) received orally (gavage) 0.2 mL of chloroquine solution 20 mg/Kg/day and control group (n=7) received filtered water by the same procedure. The animals were treated for 15 days. On the last day of treatment the mice received 0.1 mL (0.66 MBq) of Na^{99m}TcO₄⁻ via orbital plexus under anesthesia. After 40 minutes, all animals were quickly killed under anesthesia with xylazine (20 mg/kg) and ketamine (50 mg/kg), by intraperitoneal via. Kidney, liver and stomach samples were washed in 0.9% saline, and with the blood sample weighed on a precision scale (Mark 160®, Bel equipment, Italy). Percentage of radioactivity per gram of tissue (%ATI/g) was determined in an automatic gamma counter (Wizard 1470, Perkin-Elmer, Finland). The efficiency of the gamma counter was 86%, as specified by the manufacturer. The percentage of radioactivity per gram (% ATI/g) was calculated by dividing the percentage of total radioactivity of each organ by its weight in grams¹⁶. Parts of tissue samples were fixed in 10% formaline, cut as 5 µm tissue sections, stained with hematoxylin-eosin and dehydrated in ethanol and xylene for histologycal analysis¹⁸. We evaluated and quantified the changes arising on the following histological features: vascular congestion (sinusoidal, centrilobular and portal space), necrosis and fatty liver. The intensity of histopathological features was expressed in crosses (0-3 +), obtained from the average of three random microscopic fields, considering the following graduation: 0+: no change, 1+: mild changes (less than 25% of the field examined; 2+: moderate intensity changes (25 to 50% of the field examined); 3+: severe intensity changes (more than 50% of the field analyzed). All data were presented as mean ± standard deviation. The %ATI/g was compared by t-Student test and the histhological alterations were compared by Mann-Whitney test, considering p < p0.05 statistically significant. Statistica 6.0 software was used to analyze the data.

RESULTS

Table 1 shows the relationship between the controls and the treated cloroquine group. Statistically significant difference (p<0.05) in biodistribution of sodium pertechnetate (ATI%/g) was observed in blood and liverwith increased radiotracer uptake in the treated group.

Table 2 shows histological parameters in control and treated groups. Significant histological changes were seen in mesangial cells in the kidney of the treated group.

Table 3 shows histological parameters in liver in control and treated group with significant vascular congestion.

Organs -	ATI%/g		
	Control	Chloroquine	p-value
Blood	6.502 ± 0.6605	15.97 ± 1.336	0.0043*
Liver	2.845 ± 0.3439	4.708 ± 0.8044	0.0491*
Thiroyd	0.3300 ± 0.1663	1.178 ± 0.5653	0.1524
Kidney	1.217 ± 0.1538	1.652 ± 0.1267	0.0626
Stomach	4.357 ± 0.5784	3.990 ± 0.6371	0.6799
Maan CD: * n +0.05			

Table 1 – Biodistribution of $Na^{99m}TcO_4^-$ in control group and in mouse treated with cloroquine.

Mean ± SD; * *p*<0.05

Table 2 – Histological parameters of kidney in the control group and treated group with cloroquine.

Histological parameters –	Kid	n valuo	
	Control	Cloroquine	p-value
Mesangial Cells	$\textbf{2.25} \pm \textbf{0.25}$	$\textbf{3.20}\pm\textbf{0.20}$	0.019*
Lymphocytic infiltrate	$\textbf{2.29} \pm \textbf{0.18}$	2.50 ±0.22	0.533
Glomerular capillaries	$\textbf{2.28} \pm \textbf{0.28}$	$\textbf{2.40} \pm \textbf{0.40}$	0.755

Mean ± SD; * p<0.05

Table 3 – Histological parameters of liver in the control group and treated group with cloroquine

Histological parameters	Liver		
Histological parameters	Control	Cloroquine	p-value
Vascular congestion	1.1430 ± 0.1429	1.800 ± 0.2000	0.0202*
Necrosis	0.2857 ± 0.1844	0.8000 ± 0.3742	0.3434
Cell vacuolization	0.5714 ± 0.2974	1.400 ± 0.2449	0.1061
Steatosis	0.1429 ± 0.1429	0.2000 ± 0.2000	0.8763

Mean ± SD; * *p*<0.05

DISCUSSION

The pattern of a radiopharmaceutical biological behavior can be altered by interaction with natural or synthetic drugs, food, cigarettes, surgical procedures and parasitic infections and produce an unexpected result^{17,19,20,21, 22}. Knowledge of this interaction is important to avoid the misinterpretation of scintigraphic images^{16,23}. The alteration of the bioavailability of a radiopharmaceutical due to the effect of a drug in a specific tissue could aid in identifying the toxicolologic effect of a substance in an organ²⁴.

Several researches on the influence of antiparasitic drugs and radiopharmaceuticals are described in the literature. Xavier-Holanda et al. showed the Glucantime, an anti-Leishmaniadrug, increased the uptake of the radiopharmaceutical methylene disphosphonic acid, labeled with technetium-99m (^{99m}Tc-MDP) in the spleen, kidney, testicles, heart and liver of rats¹⁵. Antimalarial drugs also have shown alter the biodistribution of radiopharmaceuticals. Holanda et al.evaluated the influence of natural and synthetic antimalarial drugs (artemisinin and mefloquine, respectively) on the bioavailability of the radiopharmaceutical methylenediphosphonic acid labeled with technetium-99m (^{99m}Tc-MDP) in Wistar rats. A significant increase of %ATI occurredin spleen, liver and blood in rats treated with mefloquine. The %ATI increased significantly in artemisinin rat treated in femur, liver, lungs, spleen and blood. A significant decrease of %ATI occurred in mefloquine group in bladder, stout bowel, pancreas, kidneys, brain and also in artemisinin group in bladder, stout bowel, muscle, pancreas and kidneys²⁴.

Chloroquine is 60% bound to plasma proteins and equally cleared by the kidney and liver.In the liver, undergoes biotransformation by cytochrome p450 enzymes complex¹².To an extent of 40% to 70% chloroquine, is eliminated by renal elimination of the unchanged substance and patients with renal insufficiency may be necessary a dose reduction²⁵. Chloroquine is a weak base, accumulates in the acidic intracellular organelles, increases pH and induces osmotic swelling and permeation of acidic organelles¹⁰. However, knowledge of the mechanism of chloroquine cititoxicity is still unknown.It was reported that chloroquine could induce oxidative stress and causes oxidative disorders in cells²⁶.

The liver is the central organ of the metabolism of drugs as well as detoxification processes becomes a vulnerable organ action of chloroquine²⁷. The increased radiopharmaceutical uptake in the liver may have been a result of the metabolism of chloroquine in this organ. Metabolites and oxidative stress generated by chloroquine may have caused morphological changes in both the liver and kidneys and have altered the biodistribution of Tc-99m in the liver and blood.

Proliferation of mesangial cells is an early indicator of nephropathy progression. Diabetic nephropathy is characterized by proliferation of mesangial cells, mesangial expansion, hypertrophy and extracellular matrix accumulation²⁸. In our study we observed a significant proliferation of mesangial cells in mice treated with chloroquine. Since chloroquine may change the antioxidant status and cause certain organs become more susceptible to oxidative stress^{29,30}. Chloroquine treatment in general resulted in increase in activity of all the lysosomal enzymes in rats³¹.

Increased uptake of pertechnetate into the blood may indicate that chloroquine favors the labeling of red blood cells (RBCs) probably facilitating the passage of Tc-99m through the plasma membrane or chloroquine competes with binding sites of TC-99m in red blood cells resulting in greater availability of the radiopharmaceutical in plasma. The increased uptake of Tc-99m in the blood may be due to redox mechanisms. For this, in vitro tagging of red cells should be performed.

Chloroquine has been reported to cause damage to the kidneys, deteriorating renal function in animal models and in humans and exacerbating preexisting renal disease^{32,25}. Our results showed mesangial cell proliferation in the group of rats treated with chloroquine.The proliferation of such cells associated or not to the expansion of mesangial matrix are histologic findings frequently observed during the evolution of glomerulopathies³³. Pari e Murugan showed that treatment with chloroquine cause kidney damage including multiple focuses of hemorrhage, necrosis and swelling tubule also compared to normal kidneys and associated formation of highly reactive radicals as a consequence of oxidative stress caused by chloroquine³⁰.

Although kidney damage has occurred, no change in radiopharmaceutical uptake was verified in this organ with dose and duration of use, in this study. It was demonstrated an histological liver damage associated with high radiotracer uptake. The results may have clinical implications in patients with chronic use of chloroquine and make use of radiopharmaceuticals for scintigraphic examinations.

CONCLUSION

The use of chloroquine in mice interferes with the uptake of sodium pertechnetate in the liver and blood and causes histological changes in the liver and kidney. Although research is experimental, it is suggested clinical investigations on a possible alteration of the biodistribution of radiopharmaceuticals for scintigraphic examinations in patients using chloroquine.

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REFERENCES

- Mockenhaupt FP, May J, Bergqvist Y, Ademowo OG, Olumese PE, Falusi AG, Großterlinden L, Meyer CG, Bienzle U. Concentrations of Chloroquine and Malaria Parasites in Blood in Nigerian Children. Antimicrob Agents Chemother. 2000; 44:835-9.
- 2. Sá IM. A resistência à cloroquina e a busca de antimalariais entre as décadas de 1960 e 1980. Hist Cienc Saude Manguinhos. 2011; 18:407-30.
- 3. 3.Brasil. Ministério da Saúde. Guia prático de tratamento da malária no Brasil. Brasília: Ministério da Saúde, 2010. Disponível em: <u>http://bvsms.saude.gov.br/bvs/publicacoes/guia_pratico_malaria.pdf</u>
- 4. Bértolo MB, Brenol CV, Schainberg CG, Neubarth F, Lima FACD, Laurindo IM, et al. Atualização do consenso brasileiro no diagnóstico e tratamento da artrite reumatoide. Rev Bras Reumatol. 2007; 47:151-9.
- 5. Wallace DJ. The use of chloroquine and hydroxychloroquine for non-infectious conditions other than rheumatoid arthritis or lupus: a critical review. Lupus. 1996; 5:S59-S64.
- 6. Gupta YK, Gupta M, Aneja S, Kohli K. Current drug therapy of protozoal diarrhea. Indian J Pediatric. 2004; 71:55-8.
- Cañete R, Rivas DE, Escobedo AA, González ME, Almirall P, Brito KA.A randomized, controlled, open-label trial evaluating the efficacy and safety of chloroquine in the treatment of giardiasis in children. West Indian Med J. 2010; 59:607-11.
- 8. Lacava AC. Complicações oculares da terapêutica com a cloroquina e derivados. Arq Bras Oftalmol. 2010; 73:384-9.
- 9. 9. Teixeira RA, Borba EF, Bonfá E, Martino MF. Arrhythmias in systemic lupus erythematosus. Rev Bras Reumatol. 2010; 50:81-9.
- Shichiri M, Kono N, Shimanaka Y, Tanito M, Rotzoll DE, Yoshida Y, Hagihara Y, Tamai H, Arai H. A Novel Role for α-Tocopherol Transfer Protein (α -TTP) in Protecting against Chloroquine Toxicity. J Biol Chem. 2012; 287:2926–34.
- 11. Müller F, König J, Glaeser H, Schmidt I, Zolk O, Fromm MF, Maas R. Molecular Mechanism of Renal Tubular Secretion of the Antimalarial Drug Chloroquine. Antimicrob Agents Chemother. 2011; 55: 3091–8.
- 12. Ducharme J, Farinotti R. Clinical pharmacokinetics and metabolism of chloroquine. Focus on recent advancements. Clin Pharmacokinet. 1996; 31:257-74.
- 13. Thrall JH, Ziessman HA.Medicina Nuclear. 2^a edition. Rio de Janeiro: Guanabara Koogan;2003.
- 14. Saha GB. Fundamentals of Nuclear Pharmacy. 6^a edition. New York:Spring-Verlag;2010.
- Xavier-Holanda CMC, Jales RLC, Catanho MTJA, Holanda-Leite RC, Brito LML, Jales-Junior LH, Brandão KC, Amorim LF, Brito GGB, Gomes ML, Bernardo-Filho M. Effects of the glucantime on the kinetic of biodistribution of radiopharmaceuticals in wistar rats. Cell Mol Biol. 2002; 48:761-5.
- Bernardo-Filho M, Santos-Filho SD, Moura EG, Maiworm AI, Orlando MMC, Penas ME, et al. Drug Interaction with Radiopharmaceuticals: a Review. Braz Arch Biol Technol. 2005; 48:13-27.
- 17. Gomes ML, Oliveira MBN, Bernardo-Filho M. Drug interaction with radiopharmaceuticals: effect on the labeling of red blood cells with technetium-99m

and on the bioavailability of radiopharmaceuticals. Braz Arch Biol Technol. 2002;45:143-9.

- Tolosa EMC, Rodrigues CJ, Behmer OAF. Manual de Técnicas Para Histologia Normal e Patológica. 2ª edição. São Paulo: Monole;2003.
- Bustani H, Colavolpe C, Imbert-Joscht I, Havlik P, Pisano P, Guillet BA. Chocolate Intake Associated with Failed Labeling of ^{99m}Tc Red Blood Cells. J Nucl Med Technol. 2009; 37:107–10.
- 20. Holanda CMCX, Barbosa DA, Demeda VF, Bandeira FTB, Medeiros HCS, Pereira KRSG, Barbosa VSA, Medeiros AC .Influence of *Annona muricata* (soursop) on biodistribution of radiopharmaceuticals in rats. Acta Cir Bras. 2014; 29:145-50.
- 21. Barbosa VSA, Holanda CMCX, Câmara ACJ, Silva RP, Oliveira DP, Moreira JA, Medeiros AC. *Trypanosoma cruzi:* Biodistribution of technetium-99m pertechnetate in infected rats. Exp Parasitol. 2009;123: 309–12.
- 22. Valença SS, Lima EAC, Dire GF, Bernardo-Filho M, Porto LC. Sodium Pertechnetate (Na99mTcO4) Biodistribution in Mice Exposed to Cigarette Smoke. BMC Nucl Med. 2005; 5:1.
- Araújo-Filho I, Rego ACM, Brandão-Neto J, Villarim-Neto A, Egito EST, Azevedo IM, Medeiros AC. Biodistribution of the Radiopharmaceutical Sodium Pertechnetate after Biliopancreatic Bypass with a Duodenal Switch. Braz Arch Biol Technol.2007; 50:189-97.
- 24. Holanda CMCX, Leite RCH, Nunes RAS, Oliveira HA, Catanho MTJA, Souza GML, Bernardo-Filho M. Effect of Antimalarial Drugs on the Bioavailability of the Methylenediphosphonic Acid Labeled with Technetium-99m (^{99m}Tc-MDP) in Wistar Rats. Braz Arch Biol Technol. 2006; 49:207-14.
- 25. Müller-Hõcker J, Schmid H, Weiss M, Dendorfer U, Braun GS. Chloroquineinduced phospholipidosis of the kidney mimicking fabry's disease: Case report and review of the literature. Hum Pathol. 2003; 34:285–9.
- 26. Al-Jassabi S, Azirun M, Saad A. Biochemical studies on the role of curcumin in the potection of liver and kidney damage by anti-malaria drug, chloroquine. Am Eur J Toxicol Sci. 2011; 3:17-22.
- 27. Chaturvedi P, Mwape MP. Effect of african potato (*Hypoxis hemerocallidea*) extract on oxidative stress induced by chloroquine in albino rats. AJFAND. 2011; 11:4476-89.
- 28. Wolf G, Ziyadeh FN. Molecular mechanisms of diabetic renal hypertrophy. Kidney Int. 1999; 56: 393–405.
- 29. Toler SM, Noe D, Sharma A. Selective Enhancement of Cellular Oxidative Stress by Chloroquine: Implications for the Treatment of Glioblastoma Multiforme. Neurosurg Focus. 2006; 21:1-4
- 30. Pari L, Murugan P. Tetrahydrocurcumin: Effect on Chloroquine-Mediated Oxidative Damage in Rat Kidney. Basic Clin Pharmacol Toxicol. 2006; 99: 329–34.
- 31. Patel SP, Katewa SD; Katyare SS. Effect of antimalarials treatment on rat liver lysosomal function- an in vivo study. Indian J Clin Biochem. 2005; 20:1-8.
- 32. Murugavel P, Pari L. Attenuation of chloroquine-induced renal damage by α-lipoic acid: possible antioxidant mechanism. Ren Fail. 2004; 26:515–22.
- Pinto LMO. Revisão/Atualização em Nefrologia Clínica: Células mesangiais e matriz mesangial: sua interação mediando o processo de cronificação da lesão glomerular. J Bras Nefrol. 1998; 20:178-85.